Virus diseases of orchids in The Netherlands

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Abstract

The most commonly encountered virus diseases in orchid plants in The Netherlands are caused by *Cymbidium* mosaic virus (CyMV) and *Odontoglossum* ringspot virus (ORSV), either singly or in combination. Data on bost ranges are given.

Introduction

During the period from September 1959 until June 1960, I spent a sabbatical leave at the University of Utrecht, under the Fulbright Program, at the invitation of Professor Dr L. C. P. Kerling and as a guest in the laboratory at Baarn. She, her staff and the facilities of the Phytopathologisch Laboratorium "Willie Commelin Scholten" made possible an investigation of some of the orchid virus diseases occurring in The Netherlands.

The Canton Garden orchid collection proved to be of great value as a source of orchid species, and the commercial orchid growers in the Netherlands were generous in supplying diseased plants for experimental use.

Viruses, or virus-like symptoms, have been reported from a large number of orchid genera (Jensen, 1959). The virus diseases most commonly encountered in orchid plants in The Netherlands, as in America, were caused by two viruses either singly or in combination. These are *Cymbidium* mosaic virus (CyMV) and *Odontoglossum* ringspot virus (ORSV), the latter being related to tobacco mosaic virus. Both viruses are highly infectious, stable, and are usually present in orchid juice in high concentrations. Also both viruses have rather wide host ranges among orchid genera and species. Electron microscopy was used from time to time to confirm the identity of the respective viruses.

The present paper will discuss some data on the two important orchid virus diseases and will give a survey of my own results in studying the host ranges of CyMV and ORSV and in identifying various orchid isolated in The Netherlands.

Some data from the literature

Cymbidum mosaic

This is probably the most commonly found virus disease of orchids throughout the world. Its symptoms were described in Australia by Magee (1943), who inferred a viral etiology. Jensen (1950, 1951) experimentally transmitted the virus from *Cymbidium* to *Cymbidium*; Gold and Jensen (1951) reported electron microscopy of the causal virus

particles which are sinuous rods (about $18 \text{ nm} \times 475 \text{ nm}$), and Jensen and Gold (1955) determined that CyMV is also the cause of a common leaf necrosis disease prevalent in orchids of the genus *Cattleya* and its hybrids. It also occurs in a number of other orchid genera and can infect certain non-orchid host plants.

Odontoglossum ringspot.

This disease was described from *Odontoglossum grande* by Jensen and Gold (1951), who also reported on the size of the rod-shaped virus particles (24 nm \times 280 nm) as determined in the electron microscope. Subsequently, ORSV was shown to occur in *Cymbidium* (Jensen and Gold, 1952), where it was called *Cymbidium* diamond mottle, and in *Cattleya* (Jensen, 1959), where it causes some flower variegation.

Relationships between ORSV and TMV were evident but unclear for several years. What was obviously the same virus, based on electron microscopy, was reported from orchids by Newton and Rosberg (1952), Perez et all (1956), and Murakishi (1958b). The form and size of ORSV and TMV are so similar that they cannot always be consistently distinguished in the electron microscope (Brandes and Chessin, 1965). However, in host range and some other respects they show differences.

The most thorough investigations of the relationships of ORSV to TMV were made by Paul et al. (1965). The ORSV isolate used in their studies was one supplied by the present writer from among the *Cymbidium* plants found infected in The Netherlands (Brandes, 1964).

The conclusions reached by Paul et al. (1965) are that ORSV and TMV are distantly related serologically and are not identical. In the analytical ultracentrifuge, purified ORSV showed two peaks whereas TMV gave only one peak under identical conditions. They concluded that ORSV is related to TMV *vulgare* and that the degree of relationship is similar to that of TMV *vulgare* and Holmes' ribgrass isolate.

Inouye (1966) and Corbett (1967), on the basis of host range and serology, and Corbett (1967), from electron microscopy, also concluded that TMV and ORSV, though related, are significantly different.

Host ranges of Cymbidium mosaic and Odontoglossum ringspot viruses.

Both of these common viruses infect a wide range of orchid genera and species. Also, there is now an increasing list of non-orchid host plants which are susceptible to infection by one or both of these viruses. The viruses to which certain of these plants are susceptible, however, have not been unequivocally determined. It is probable that in some cases both viruses, instead of only one, were present in the inoculum or that the single virus used was misidentified.

Datura stramonium was shown by White and Goodchild (1955) to develop local lesions when inoculated with CyMV. It is apparently not susceptible to ORSV. An even better indicator for the presence of CyMV (and the absence of ORSV) is Cassia occidentalis, which develops local lesions in 4–6 days (Corbett, 1960), as do also C. tora and C. bacapsularis. Also reported as hosts of CyMV are Tropeolum majus L., Oryza sativa L., Passiflora edulis Sims and Zinnia elegans Jacq. (Murakishi, 1958a).

Gomphrena globosa is an excellent selective indicator host plant for ORSV which causes local lesions on the inoculated leaves in approximately 10 days but does not react to CyMV. Thus, a mixture of CyMV and ORSV can be separated and identified by inoculation to Cassia and Gomphrena.

In an extensive and valuable host range study, Hollings (1959) tested 52 viruses on 15 different non-orchid plant genera. One of the viruses was listed as *Cymbidium* mosaic virus derived from *Cymbidium*. However, the inoculum used in the host range experiment came from *Gomphrena* or *Primula*. Thirteen of the fifteen genera and species inoculated were recorded as positive.

We now know that Gomphrena globosa is not a host of CyMV, but is susceptible to ORSV. It is probable, therefore, that Hollings' inoculum from Gomphrena was actually ORSV rather than CyMV. Other plants reported by Hollings as being susceptible to the orchid virus or viruses in his inoculum were: Chenopodium amaranticolor Coste & Reyn., Beta vulgaris L., Amaranthus caudatus L., Celosia argentea L., Tetragonia expansa Murr., Phytolacca americana L., Portulaca oleracea L., Fagopyrum esculentum Moench., Primula malacoides Franch., Lythrum salicaria L., and Stellaria media Vill. The latter five were listed as symptomless. Only Primula and Stellaria were systemically invaded.

Host range studies

The writer's orchid investigations in The Netherlands included host range studies of orchid viruses involving some of the above test plant species used by Hollings. On the basis of a large number of inoculations from many orchid virus sources, the following conclusions were drawn. Cymbidium mosaic virus produced local lesions on Datura stramonium and Cassia occidentalis but caused no symptoms in Tetragonia, Gomphrena Chenopodium amaranticolor, Nicotiana tabacum 'Xanthi', N. tabacum 'Samsum', or Zinnia elegans.

Odontoglossum ringspot virus caused local lesions on Gomphrena, Chenopodium, Nicotiana tabacum 'Xanthi' and Tetragonia, but caused no symptoms on Cassia, Datura, or Amaranthus.

Amaranthus plants were inoculated with CyMV from four sources, with ORSV from six sources and with a mixture of both viruses from four sources. In no case were there decisive symptoms of infection.

Chenopodium developed local lesions after inoculation from eight sources of ORSV only, and from four sources carrying both ORSV and CyMV, but in none of seven experiments in which the inoculum was CyMV only.

Among the additionally known hosts of ORSV are Zinnia elegans (Jensen and Gold, 1952), in which the only symptoms was a transitory variegation of the flowers, Chenopodium quinoa (Brandes, 1964), C. capitatum and C. ficifolium van Regenmortel et al., 1964).

Nicotiana tabacum 'Samsum' was found by Paul et al. (1965) to develop no symptoms of ORSV infection, but the virus reached a high concentration in the inoculated leaves after a build up of 3 weeks. They also found that N. tabacum 'Xanthi' was a good local lesion host in which ORSV did not become systemic. Although Nicotiana glutinosa is a good local lesion host for TMV, most efforts to use it as an indicator for ORSV have been unsuccessful. However, Paul et al. (1965) found that although ORSV produced no symptoms in this plant in the summer, inoculated leaves did develop grey flecks of variable size in the spring and fall.

Identification of Dutch isolates

In addition to Cymbidium, Cattleya and Cattleya hybrids, orchid viruses were recover-

ed from other orchid genera in The Netherlands. Calanthe, with chlorotic spots on the leaves, yielded inoculum that produced CyMV symptoms in Cymbidium, Cassia and Datura. Lesions were also produced in Chenopodium but not in Gomphrena. Laelia rubescens, with mosaic leaf symptoms, carried CyMV but not ORSV. Vanda sumatrana, with black lesions on the leaves, yielded juice that produced virus symptoms in Spathoglottis orchid but failed to infect Cymbidium or the indicator hosts for either CyMV or ORSV.

Miltonia regnelli Rchb.f. had mosaic leaf symptoms which may have been due to CyMV present in very low concentration, as suggested by a very few lesions on the inoculated cotyledons of Cassia.

Ada aurantiaca, with numerous brown to rose-colored spots on the under surface of the leaves, and Schomburgkia crispa Lindl., with mosaic leaves, were found to be infected with CyMV.

A *Phalaenopsis* plant, with white sunken areas on the older leaves, was carrying both CyMV and ORSV.

Variegated flowers were found, but only rarely, on Cymbidium plants in The Netherlands. They resembled somewhat the color breaking of Cymbidium flowers in Japan illustrated by Inouye (1965). The identity of the virus in The Netherlands is still obscure because when juice from the flowers was inoculated to Nicotiana tabacum 'Xanthi', Cassia, Gomphrena, Tetragonia and Phytolacca americana, symptoms occurred only on N. xanthi, where numerous lesions developed on the inoculated leaves.

Samenvatting

Virusziekten van orchideeën in Nederland

De meest algemeen voorkomende virusziekten van orchideeën in Nederland worden veroorzaakt door *Cymbidium*-mozaïekvirus (CyMV) of *Odontoglossum*-kringvlekkenvirus (ORSV) of door een combinatie van beide virussen. Aan de waardplantenreeks van beide virussen wordt uitvoerig aandacht geschonken.

References

Brandes, J., 1964. Identifizierung von gestreckten pflanzenpathogenen Viren auf morphologischer Grundlage. Mitt. biol. Bund. Anst. Ld- u. Forstw. 110.

Brandes, J. & Chessin, M., 1965. An electron microscope study on the size of Sammons' Opuntia virus. Virology 25: 673-674.

Corbett, M. K., 1960. Purification by density gradient centrifugation, electron microscopy, and properties of Cymbidium mosaic virus. Phytopathology 50: 346–351.

Corbett, M. K., 1967. Some distinguishing characteristics of the orchid strain of tobacco mosaic virus. Phytopathology 57: 164-172.

Gold, A. H. & Jensen, D. D., 1951. An electron microscope study of Cymbidium mosaic virus. Am. J. Bot. 38: 577-578.

Hollings, M., 1959. Host range studies with fifty-two plant viruses. Ann. appl. Biol. 47: 98-108.

Inouye, N., 1965. Virus diseases of orchids. II. Symptoms and properties of viruses in Cymbidium. Japan Orchid Soc. Bull. 11: 1-6.

Inouye, N., 1966. A virus disease of Cymbidium caused by Odontoglossum ringspot virus. Ber. Ohara Inst. Landw. Biol. 13: 149-159.

Jensen, D. D., 1950. Mosaic of Cymbidium orchids. Phytopathology 40: 966-967.

Jensen, D. D., 1951. Mosaic or black streak disease of Cymbidium orchids. Phytopathology 41: 401-414

- Jensen, D. D., 1952. Virus diseases of orchids. Calif. Agric. 6: 7, 15, 16.
- Jensen, D. D., 1959. Virus diseases of orchids. In: The Orchids, p. 431-458. The Ronald Press Co.
- Jensen, D. D. & Gold, A. H., 1951. A virus ringspot of Odontoglossum orchid: symptoms, transmission, and electron microscopy. Phytopathology 41: 648-653.
- Jensen, D. D. & Gold, A. H., 1952. Virus diseases of orchids in California. Phytopathology 42: 12.
- Jensen, D. D. & Gold, A. H., 1955. Hosts, transmission and electron microscopy of Cymbidium mosaic virus with special reference to Cattleya leaf necrosis. Phytopathology 45: 327–334.
- Magee, C. J., 1943. Orchid mosaic. Aust. Orchid Rev. 8: 51-52.
- Murakishi, H. H., 1958a. Host range, symptomatology, physical properties, and cross-protection studies of orchid virus isolates, Phytopathology 48: 132–137.
- Murakishi, H. H., 1958b. Serological and morphological relationships among orchid viruses. Phytopathology 48: 137–140.
- Newton, N. & Rosberg, D. W., 1952. Electron-microscope studies of a new orchid virus complex. Phytopathology 42: 79–82.
- Paul, H. L., Wetter, C., Wittmann, H. G. & Brandes, J., 1965. Untersuchungen am Odontoglossum ringspot virus, einem Verwandten des Tabakmosaik-Virus. Z. Vererblehre 97: 186–203.
- Perez, J. E., Adsuar, J. & Sala, O. 1956. Tobacco mosaic virus in orchids in Puerto Rico. Phytopathology 46: 650-654.
- Regenmortel, M. H. V. van, Hahn, J. S., and Fowle, L. G., 1964. Internal calibration of electron micrographs with an orchid virus. S. Afr. J. agric. Sci. 7: 159–164.
- White, N. H. & Goodchild, D. J., 1955. Mosaic or black streak disease of Cymbidium and other orchid hybrids. J. Aust. Inst. agric. Sci. 21: 36-38.